Body Fluid Identification by Proteomic Mass Spectrometry - Liquid Chromatography & Mass Spectrometer Processing

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Body Fluid Identification by Proteomic Mass Spectrometry -Liquid Chromatography & Mass Spectrometer Processing

1 Purpose

1.1 Samples are run through liquid chromatography to separate peptide markers. Marker peptides are identified by mass spectrometry.

2 Check Before Starting

2.1 Check the Pure Air tank. If pressure on the large gauge (right gauge) is \leq 300 psi, <u>do not</u> <u>proceed</u>. Contact the Proteomics Team to change the Pure Air tank if it is less than 300 psi.



- 2.2 Check the LC/MS instrument to see how many batches are in the queue. Ensure reagents are enough for batches in queue.
- 2.3 Check all volumes in reagent bottles. If reagent bottles do not have the volumes listed below, **do not proceed**. Contact the Proteomics Team to change the reagents bottles if their volumes are less than the volumes listed below.
 - 2.3.1 Methanol (50% methanol + 50% HPLC water) 200 mL
 - 2.3.2 Loading Solution (phase A 2% ACN + 98% HPLC water) 200 mL
 - 2.3.3 Bottle A (HPLC water) 160 mL

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- 2.3.4 Bottle B (ACN) 120 mL
- 2.4 For **samples that need to be rerun**, follow the instructions above (2.1 to 2.3 above) and then proceed to section 3.9.

3 Liquid Chromatography – Mass Spectrometer Analysis Procedure

3.1 Retrieve the following reagents:

Cytochrome C (8)	<mark>pmol/ μl)</mark> at -80°C	
PCM Standard		
Phase A		
Acetone		
Acetonitrile		
Isopropanol		

- 3.2 Prepare a working solution of Cytochrome C Master Mix sufficient to resuspend all samples (see 3.3) Work on ice or in a -20°C cold tube rack.
- 3.3 Input into LIMS the number of regular quant samples + the number of low quant samples divided by eight. If the number is a fraction round up to the next integer. For example, if you have five regular quants and three low quant samples you have 5 + 3/8 = 5.38, therefore input 6 into LIMS.

Cytochrome C Master Mix Reagent Ratios (multiply volumes by the number of samples to be resuspended)				
Reagent	Volume			
Phase A	<mark>93 µl</mark>			
Cytochrome C <mark>(8 pmol/µl)</mark>	<mark>1.45 μl</mark>			
Total volume:	<mark>94.45 μl</mark>			

- 3.4 Resuspend samples as follows:
- 3.5 **REGULAR Samples**: add 94.45µl of Cytochrome C Master Mix (8 pmol/µl) to reconstitute peptides.
- 3.6 **LOW Concentration Samples**: add 10 μl of Cytochrome C Master Mix solution to reconstitute peptides.
- 3.7 Vortex.
- 3.8 Place in refrigerated centrifuge at 4°C and spin at 18,000 g (RCF) for 30 minutes. Record instrument and temperature in LIMS.

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- 3.9 Pipette supernatant into LC Vial (avoid pellet if present) and store at 4°C.
- 3.10 Prepare the LC cleaning solution to be run alongside samples and controls:

LC Cleaning Solution				
Reagent	Volume			
Acetone	2 µl			
Acetonitrile	9 μ1			
Isopropanol	9 μl			
Total Volume:	20 µl /sample			

- 3.11 Place samples, controls, and prepared cleaning solution in Eksigent refrigerated auto sampler and fill out location of each vial in the input load plate on LIMS.
- 3.12 Fill in LIMS ouput data entry sheet and select the appropriate Acquistion Method for the sample type.

Sample Type	AcqMethod
PepCalMix, Phase A	Col1 PepCalMix Col2 PepCalMix
Cleaning Sol	Col1 Cleaning Col2 Cleaning
ENeg, Ext Pos controls, Unknowns	Col1 Unknown Col2 Unknown
Reruns (High)	Col1 Unknown Hi Col2 Unknown Hi

3.12.1 Your batch should not end on a "col 1" method. If your batch does end in a "col 1" method, add an additional phase A line at the end of the batch with a "col 2" method.

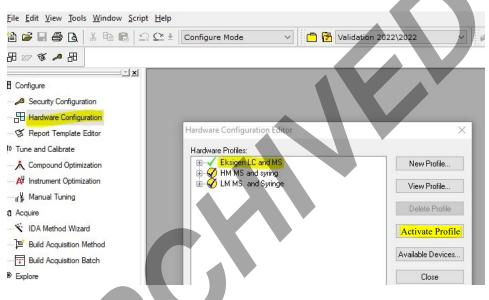
- 3.13 Output Plate Name should be the date (YYYYMMDD), followed by a letter identifying batch order (e.g. A, B, C, etc.) an underscore, followed by analyst initials.
- 3.14 Ensure the output samples are in the desired injection order before loading plate (in its default vertical mode)
 - 3.14.1 Positive controls should be run after unknown samples when creating batch.
- 3.15 Export sample batch from the output plate in LIMS.
- 3.16 Open LIMS created batch excel sheet in the LIMS LCMS folder and confirm it is as desired and save document as .txt format.

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- 3.17 Open analyst software.
- 3.18 Double click Hardware Configuration \rightarrow Eksigent LC and MS \rightarrow Activate Profile



3.19 Ensure that Eksigent windows is open simultaneously (both Eksigent Control Software window and the Eksigent Autosampler window).

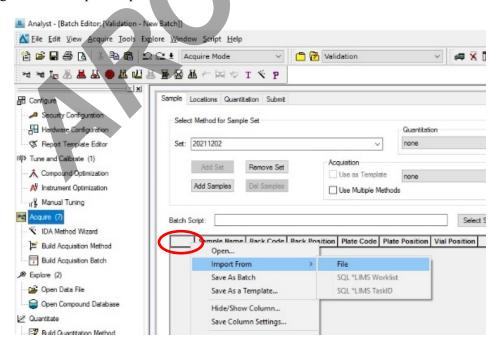
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3.20 Double click Build Acquisition Batch

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Right click to import acquisition batch. 3.21



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3.22 Select .txt batch list exported from LIMS. Click open.

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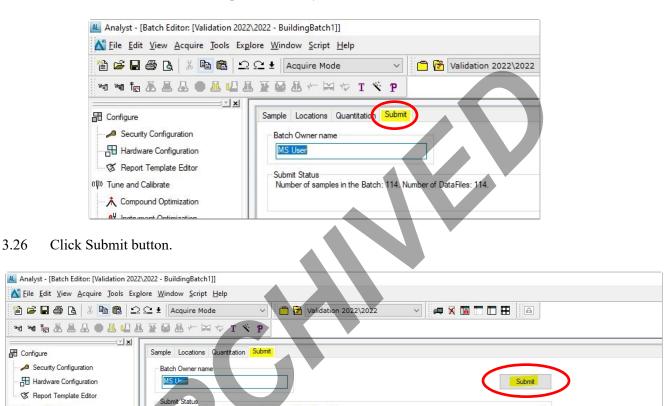
3.23 Select autosampler (ekspert nanoLC 400). Click OK.

Select Autosampler		
Alias CTC PAL	^	OK
CTC-MPX Driver		Cancel
Dionex ACC-3000		Carica
Dionex AS(50) Dionex ASI-100		
Dionex WPS-3000		
Eksigent AS1		
ekspert nanoLC 400 Endurance		

3.24 **Label and Position WITNESS**: Have a witness verify the selected autosampler, batch sample names, methods, and tube positions in autosampler match that in Vial Position column.

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3.25 Click on Submit tab when sample list is ready.



3.27 Ensure "All samples" is selected and that the box for "apply new samples order" is unchecked. Click OK

Number of samples in the Batch: 114. Number of DataFiles: 114.

Acquisition	>
Acquire data for:	
All samples	
Apply new samples order	
OK Cancel	Help

(I)) Tune and Calibrate

aU

A Compound Optimization

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3.28 Click Queue button and double check all samples were submitted.

Analyst - [Queue Manager [Local]]										
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3.29 Click Ready button. Analyst - [Queue Manager [Local]] Eile Edit View Acquire Tools Explo							- 147		_	
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3.30 Click Start sample button. The Eksigent LC and 6500 MS will process all samples.

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